

## Role of calcium and calmodulin antagonist in photosynthesis and salinity tolerance in *Chlorella vulgaris*

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### Abstract

To cast light upon the role of  $\text{Ca}^{2+}$  and calmodulin on photosynthetic rate ( $P_N$ ), dark respiration ( $R_D$ ) and amino acid and protein contents in salinity stressed and non-stressed *Chlorella* cultures, the  $\text{Ca}^{2+}$  chelator EGTA [ethylene glycol-bis-(2-aminoethyl ether)-*N,N*-tetraacetate] and the calmodulin antagonist TFP (trifluoperazine) were used. TFP markedly inhibited  $P_N$  while EGTA exerted a slight, if any, effect on  $P_N$ . NaCl tolerance, on the other side, was markedly abolished by TFP that inhibited  $P_N$  and lowered rate of proline accumulation. Calmodulin might be involved in osmoregulation and salt tolerance of *Chlorella*.  $R_D$ , however, was markedly enhanced by EGTA and  $\text{Ca}^{2+}$ -free medium and hence the  $\text{Ca}^{2+}$  deprivation increased stress severity exerted by NaCl. Combinations of  $\text{Na}^+$  and  $\text{Ca}^{2+}$  enhanced  $P_N$ , decreased  $R_D$  and proline content in comparison with an osmotically equivalent reference culture containing only NaCl. Addition of  $\text{Ca}^{2+}$  to TFP treated cultures failed to reactivate calmodulin for proline synthesis. However, when  $\text{Ca}^{2+}$  was added to EGTA-treated cultures, only relatively reduced proline contents were recorded.

### Introduction

The regulation of calcium level is a major mechanism of plant cell metabolism.  $\text{Ca}^{2+}$  has a role in the maintenance of membrane integrity (Leopold and Willing 1984), cell elongation (Pickard 1970), amylase secretion (Jones and Jacobsen 1983), delay of senescence (Ferguson 1984), and several other processes. In addition,  $\text{Ca}^{2+}$  regulates various biochemical processes after binding to calmodulin and related proteins (Cheung 1980). Photosynthesis and respiration has also been correlated with  $\text{Ca}^{2+}$  and calmodulin (Bangerth *et al.* 1972, Woodrow and Rowan 1979, Jarett *et al.* 1982, Muto *et al.* 1982, Burris *et al.* 1983, Ferguson 1984).  $\text{Ca}^{2+}$  may be involved in the conformational changes in the thylakoids and in the coupling of electron flow (Gross *et al.* 1976). In addition,  $\text{Ca}^{2+}$  is an important factor in the resistance of plants to salinity (Lahye and Epstein 1971, Greenway and Munns 1980, Kent and Läuchli

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1985). Abdel-Basset (1986) found that  $\text{Ca}^{2+}$  in certain ratios to  $\text{Na}^+$  ( $\text{Na}^+/\text{Ca}^{2+}$  of 13.5) reversed most of NaCl stress symptoms in *Chlorella vulgaris*. This ratio improved the growth and photosynthetic rate ( $P_N$ ), and lowered both the respiration rate ( $R_D$ ) and proline accumulation rates of salinized *Chlorella* cultures (stress relief).

The aim of this work was to follow the response of *Chlorella vulgaris* to NaCl in calmodulin inhibited cultures (using TFP) or in  $\text{Ca}^{2+}$ -chelated cultures (using EGTA). In addition, the effects of isoosmotic  $\text{Na}^+/\text{Ca}^{2+}$  combinations were investigated.  $P_N$ ,  $R_D$  and protein, proline and other free amino acids contents were analysed in short-term experiments.

### Material and methods

Unialgal cultures of *Chlorella vulgaris* Beijerinck (isolated from the river Nile) were grown in batch cultures using the Beijerinck's nutritive medium (Stein 1966) under continuous irradiation ( $5.9 \text{ W m}^{-2}$ ) and aeration at temperature  $25 \pm 1^\circ\text{C}$ . This medium contains  $3.6 \text{ g}(\text{Ca}^{2+}) \text{ m}^{-3}$  and  $103 \text{ g}(\text{P}) \text{ m}^{-3}$ . The aliquots of algal suspension were studied without dilution and at the same conditions of growth under the following treatments:

EGTA [ethylene glycol-bis(2-aminoethyl ether)-N,N-tetraacetate (Serva, Heidelberg, Germany)] : 2, 4 or 7 mM was applied to check the responsiveness of  $P_N$  and  $R_D$  to  $\text{Ca}^{2+}$  chelation.

$\text{Ca}^{2+}$  free medium: Algal cells were washed twice in the Beijerinck's medium free of  $\text{Ca}^{2+}$  and were allowed to grow for further 48 h to determine the effects of  $\text{Ca}^{2+}$  deprivation (or at least dilution).

TFP [trifluoperazine, (Fluka, Buchs, Germany)]: 20, 40 and 100  $\mu\text{M}$  were applied to check the responsiveness of  $P_N$  and  $R_D$  to calmodulin antagonists.

Salinization: 100, 200 and 300 mM NaCl were applied. To clarify the role of  $\text{Ca}^{2+}$  and calmodulin with respect to salt tolerance EGTA (7 mM) or TFP (0.1 mM) was added to variously salinized cultures of *Chlorella*.

$\text{Na}^+/\text{Ca}^{2+}$  combinations: Isoosmotic combinations with  $\text{Na}^+/\text{Ca}^{2+}$  of 5, 10 and 15 were used (Ca as  $\text{CaCl}_2$ ). These were osmotically equivalent to the moderate salinization (reference culture contained only 200 mM NaCl). The  $\text{Na}^+/\text{Ca}^{2+}$  combinations were also applied in the presence of TFP to understand the role of calmodulin.

$P_N$  and  $R_D$  were measured under the various EGTA or TFP concentrations immediately after their addition. In salinized cultures, the assay and analysis were measured after 3 h (short-term experiments) with NaCl,  $\text{CaCl}_2$ , EGTA and TFP. The mean values of three replicates were presented.  $P_N$  (oxygen evolution) and  $R_D$  (oxygen uptake in darkness) were determined polarographically using an oxygen electrode (Schott Geräte GmbH, Germany) under the same irradiance as the growth ( $5.9 \text{ W m}^{-2}$ ). The chlorophyll content was determined spectrophotometrically in acetone extracts using the equations recommended by Metzner *et al.* (1965).

The proline was extracted by sulphosalicylic acid and then determined colorimetrically after Bates *et al.* (1973). The total free amino acids were estimated by the Lee and Takahashi (1966) method. The soluble proteins were determined in the boiling water extracts according to the procedure described by Lowry *et al.* (1951).

## Results

$\text{Ca}^{2+}$  free medium reduced  $P_N$  of *C. vulgaris* (Table 1). EGTA reduced  $P_N$  only at the highest concentration (7 mM), but lower concentrations of EGTA enhanced it (Table 1). However, at much lower concentrations than EGTA, TFP markedly inhibited  $P_N$  as soon as it was added.

Table 1. Photosynthetic oxygen evolution ( $P_N$ ) and respiratory oxygen consumption ( $R_D$ ) of *Chlorella vulgaris* cultures as influenced by EGTA or TFP immediately after addition expressed as percentage of the control (without salinity).  $P_N$  and  $R_D$  of the control culture were 15.8 and 1.9 mmol( $\text{O}_2$ )  $\text{kg}^{-1}(\text{Chl}) \text{ s}^{-1}$ , respectively.

| Treatment             | $P_N$ | $R_D$ | $P_N/R_D$ |
|-----------------------|-------|-------|-----------|
| control               | 100   | 100   | 9.1       |
| -Ca                   | 67    | 171   | 4.2       |
| EGTA 2 mM             | 112   | 129   | 8.1       |
| EGTA 4 mM             | 112   | 171   | 6.3       |
| EGTA 7 mM             | 88    | 157   | 5.6       |
| TFP 20 $\mu\text{M}$  | 56    | 86    | 6.3       |
| TFP 40 $\mu\text{M}$  | 56    | 200   | 3.3       |
| TFP 100 $\mu\text{M}$ | 46    | 243   | 2.5       |

After being salinized for 3 h,  $P_N$  was lowered more at a higher NaCl concentration (Table 2). TFP treated cultures exhibited a markedly inhibited  $P_N$  with a net  $\text{O}_2$  uptake occurring in the light. However, EGTA enhanced  $P_N$  of variously NaCl-salinized *Chlorella* cultures.

At a  $\text{Na}^+/\text{Ca}^{2+}$  ratio of 10,  $P_N$  was higher in comparison with the isoosmotic reference culture containing only NaCl. However, TFP treated cultures exhibited a markedly lowered  $P_N$  in the presence of these combinations (Table 2).

$R_D$  was affected by all treatments.  $\text{Ca}^{2+}$ -free medium, EGTA and TFP caused an enhanced  $R_D$  (Table 1) immediately after addition. NaCl also enhanced  $R_D$  after 3 h. Chelation of  $\text{Ca}^{2+}$  by EGTA in salinized cultures resulted in higher  $R_D$  than in the control or salinized cultures not treated with EGTA. However, TFP markedly lowered  $R_D$  of salinized cultures (Table 2). The combinations of  $\text{Na}^+$  and  $\text{Ca}^{2+}$  lowered  $R_D$  compared with the control (Table 2). However, TFP markedly elevated  $R_D$  of the cultures treated with these combinations.

The  $P_N/R_D$  ratio was lowered with increasing NaCl level, while it increased at  $Na^+/Ca^{2+}$  of 10. TFP markedly lowered this ratio to minimal values.

Table 2. Photosynthetic oxygen evolution ( $P_N$ ) and respiratory oxygen consumption ( $R_D$ ) of *Chlorella vulgaris* cultures as influenced by salinization treatments (NaCl or NaCl +  $CaCl_2$ ) in the presence of 7 mM EGTA or 100  $\mu$ M TFP expressed as percentage of the control (without salinity).  $P_N$  and  $R_D$  of the control culture were 15.8 and 2.2 mmol( $O_2$ )  $kg^{-1}$ (Chl)  $s^{-1}$ , respectively.

| Treatment                 | $P_N$ | $R_D$ | $P_N/R_D$ |
|---------------------------|-------|-------|-----------|
| control                   | 100   | 100   | 8.8       |
| 100 mM NaCl               | 69    | 100   | 6.4       |
| 100 mM NaCl + EGTA        | 159   | 83    | 16.6      |
| 100 mM NaCl + TFP         | 0     | 43    | 1.0       |
| 200 mM NaCl               | 65    | 113   | 5.4       |
| 200 mM NaCl+ EGTA         | 133   | 150   | 8.2       |
| 200 mM NaCl + TFP         | -3    | 57    | 0.5       |
| 300 mM NaCl               | 58    | 125   | 4.6       |
| 300 mM NaCl + EGTA        | 133   | 167   | 7.5       |
| 300 mM NaCl + TFP         | -5    | 57    | 0.3       |
| $Na^+/Ca^{2+} = 5$        | 53    | 75    | 6.5       |
| $Na^+/Ca^{2+} = 5$ + TFP  | 8     | 186   | 1.4       |
| $Na^+/Ca^{2+} = 10$       | 95    | 50    | 15.8      |
| $Na^+/Ca^{2+} = 10$ + TFP | -9    | 186   | 0.5       |
| $Na^+/Ca^{2+} = 15$       | 32    | 63    | 5.0       |
| $Na^+/Ca^{2+} = 15$ + TFP | -20   | 286   | 0.4       |

Table 3. Contents of proline, other free amino acids and soluble and insoluble proteins related to chlorophyll content [ $g\ kg^{-1}$ (Chl)] as influenced by various salinization treatments in the presence of 7 mM EGTA or 100  $\mu$ M TFP.

| Treatment                 | Proline | Amino acids | Soluble proteins | Insoluble proteins | Total proteins | Ratio sol./insol. |
|---------------------------|---------|-------------|------------------|--------------------|----------------|-------------------|
| control                   | 35      | 109         | 1717             | 3902               | 5619           | 0.44              |
| 100 mM NaCl               | 39      | 57          | 1172             | 2930               | 4102           | 0.40              |
| 100 mM NaCl + EGTA        | 89      | 53          | 1663             | 2148               | 3811           | 0.77              |
| 100 mM NaCl + TFP         | 24      | 66          | 747              | 5078               | 5825           | 0.13              |
| 200 mM NaCl               | 80      | 61          | 1235             | 3125               | 4360           | 0.40              |
| 200 mM NaCl+ EGTA         | 107     | 67          | 1484             | 2474               | 3958           | 0.60              |
| 200 mM NaCl + TFP         | 33      | 72          | 619              | 4688               | 5307           | 0.13              |
| 300 mM NaCl               | 59      | 65          | 1953             | 4102               | 6055           | 0.48              |
| 300 mM NaCl + EGTA        | 113     | 80          | 1328             | 3051               | 4379           | 0.44              |
| 300 mM NaCl + TFP         | 29      | 72          | 765              | 4427               | 5192           | 0.17              |
| $Na^+/Ca^{2+} = 5$        | 58      | 73          | 931              | 4011               | 4942           | 0.23              |
| $Na^+/Ca^{2+} = 5$ + TFP  | 31      | 109         | 322              | 4688               | 5010           | 0.07              |
| $Na^+/Ca^{2+} = 10$       | 53      | 81          | 846              | 3646               | 4492           | 0.23              |
| $Na^+/Ca^{2+} = 10$ + TFP | 31      | 121         | 699              | 2865               | 3564           | 0.24              |
| $Na^+/Ca^{2+} = 15$       | 50      | 85          | 804              | 3464               | 4268           | 0.23              |
| $Na^+/Ca^{2+} = 15$ + TFP | 23      | 59          | 547              | 4818               | 5365           | 0.11              |

Proline was markedly accumulated in the NaCl-salinized *Chlorella* cultures. More or less the same trend but with higher contents of proline was found in the salinized cultures treated with EGTA (Table 3). However, the NaCl in combination with  $\text{CaCl}_2$  lowered the proline contents compared with that ones in the isoosmotic reference culture containing only 200 mM NaCl. TFP markedly lowered the proline accumulation (Table 3).

The contents of other free amino acids were generally lowered by NaCl. Both EGTA and TFP reduced the amino acid contents of salinized cultures. However, relatively high concentrations of amino acids were induced by  $\text{Na}^+$ - $\text{Ca}^{2+}$  combinations.

Similarly, the contents of total protein was variably lowered under most of the treatments used (Table 3). A drastic lowering of the protein concentration was observed in EGTA treated cultures under NaCl, while the soluble/insoluble protein ratios were highest in these cultures. However, the combinations of NaCl and  $\text{CaCl}_2$  did not exert drastic changes in the protein content compared with those of the corresponding isoosmotic reference culture containing only 200 mM NaCl.

## Discussion

$P_N$  of *C. vulgaris* cultures was reduced in  $\text{Ca}^{2+}$ -free medium or by the highest EGTA concentration used (7 mM). At much lower concentrations than EGTA, TFP markedly inhibited  $P_N$  as soon as it was added. Similar results were obtained by Barr *et al.* (1982) and Burris *et al.* (1983) who agreed that TFP markedly inhibited photosynthesis. The inhibition by TFP implies the involvement of calmodulin in photosynthesis (Barr *et al.* 1982). A calmodulin-like protein was found in the stroma of pea chloroplasts (Jarret *et al.* 1982) and a Ca-calmodulin enzyme may be involved in the activation of chloroplast enzymes (Muto *et al.* 1982).  $\text{Ca}^{2+}$  ions may be involved in a photosynthesis through the a spillover phenomenon (Gross *et al.* 1976), in the conformational changes in the thylakoids and in the coupling of electron flow (Barber 1976).

After being salinized for 3 h,  $P_N$  of *C. vulgaris* was lowered and an addition of TFP entirely inhibited it. This reveals that photosynthesis of calmodulin-inhibited *Chlorella* cells is remarkably sensitive to NaCl.

The  $\text{Na}^+/\text{Ca}^{2+}$  ratio of 10 improved  $P_N$  compared with that one of the isoosmotic reference culture containing only NaCl. Thus  $\text{Ca}^{2+}$  may probably repair the NaCl injured thylakoids. A possibility is that  $\text{Ca}^{2+}$  reduces the  $\text{Na}^+$  uptake and thus its toxicity is diminished. The TFP treated cultures had a low  $P_N$ . However, Barr *et al.* (1983), working with non-salinized cultures, found that the addition of  $\text{Ca}^{2+}$  to the TFP-treated cultures relatively protected the spinach chloroplasts and stimulated the electron transport.

On the other hand,  $R_D$  was enhanced in  $\text{Ca}^{2+}$ -free medium by EGTA and TFP immediately and in salinized cultures after 3 h. Higher  $R_D$  was observed in the salinized cultures treated with EGTA. However, TFP markedly lowered the  $R_D$  of salinized cultures. In this respect,  $\text{Ca}^{2+}$  deficiency enhanced  $R_D$  (Ferguson 1984)

while high  $\text{Ca}^{2+}$  contents reduced it (Bangerth *et al.* 1972). A high  $R_D$  in  $\text{Ca}^{2+}$ -chelated cultures may indicate that the stress severity of NaCl is elevated. Consequently, a higher energy demand for osmoregulation, repair of injured tissues and active ion uptake (Gale 1975, Schwarz and Gale 1981, Lambers *et al.* 1983) is required. The  $R_D$  rise in  $\text{Ca}^{2+}$ -deficient cells is probably due to the loss of membrane stability (Ferguson 1984) that includes a breakdown of compartmentation and an increase in the contents of respiratory substrates from the vacuole to the cytoplasm; this finally enhances  $R_D$  (Woodrow and Rowan 1972, Ferguson 1984). These processes can be retarded by  $\text{Ca}^{2+}$  (Bangerth *et al.* 1972). In salinized plants,  $\text{Na}^+$  displaces  $\text{Ca}^{2+}$  from membranes (Lynch and Läuchli 1988) and the loss of membrane stability occurs probably because of  $\text{Ca}^{2+}$  deficiency. The combinations of  $\text{Na}^+$  and  $\text{Ca}^{2+}$  resulted in a remarkably lower  $R_D$  than in the control cultures. However, TFP elevated the  $R_D$  of the *Chlorella* cultures subjected to  $\text{Na}^+$ - $\text{Ca}^{2+}$  combinations.

Since the nitrogenous compounds are common factors in osmoregulation, the contents of proline, free amino acids and proteins were considered in this work. Proline was accumulated in the *Chlorella* cultures salinized for 3 h and especially in the cultures treated with EGTA. The chelation of intracellular  $\text{Ca}^{2+}$  thus elevated the stress severity exerted by NaCl, as in the case of elevated  $R_D$ . The physiological significance of proline accumulation may be the osmoregulation (Barnett and Naylor 1966), the provision of both carbon and nitrogen for the post-stress relief (Thompson *et al.* 1966), or the removal of ammonia (Henkel 1966). The  $\text{Na}^+$ - $\text{Ca}^{2+}$  combinations lowered the proline contents compared with the isoosmotic reference culture. In this respect, Dreier (1983) and Imamul Huq and Lahrer (1984) found that  $\text{Ca}^{2+}$  lowered the proline levels of stressed plants. Since the proline accumulation is simultaneously indicative for the stress damage (Stewart and Hanson 1980),  $\text{Ca}^{2+}$  may play an important role in the NaCl stress counteraction. On the other hand, the inhibition of proline accumulation under stress by TFP may indicate that calmodulin is involved in the synthesis of proline.

The contents of free amino acids and proteins were lowered under various treatments. A drastic lowering of the total protein content was observed in the presence of EGTA while the soluble/insoluble protein ratios were the highest ones. High ratios of sol./insol. proteins may be due to an increased proteolytic activity.

Generally, salt tolerance of *C. vulgaris* might involve  $\text{Ca}^{2+}$  and calmodulin. The presence of EGTA elevated proline contents and enhanced  $R_D$ . TFP turned *Chlorella* cells less salt tolerant. Nevertheless,  $\text{Ca}^{2+}$  could not recover the activity of calmodulin in the TFP-treated *Chlorella* cultures.

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