

Table 1. Suppl. A summary of spectrophotometric estimation of total RNA isolated from three different biological replicates from few representative tissues used in this study.

Species name	Tissue	Biological replicates	Concentration [$\mu\text{g cm}^{-3}$]	A _{260/280}
<i>Bambusa balcooa</i>	Young leaf	1	596.3	1.87
		2	635.1	1.96
		3	861.7	2.03
<i>B. bambos</i>	Internode	1	746.7	2.01
		2	625.6	1.96
		3	567.3	1.92
<i>B. vulgaris</i>	Root	1	634.5	1.95
		2	587.8	1.90
		3	686.8	2.01
<i>B. tulda</i>	Cum sheath	1	2106.2	2.05
		2	749.6	2.01
		3	836.3	2.02
<i>B. tulda</i>	Early staged inflorescence bud	1	435.3	1.95
		2	524.0	1.91
		3	517.3	1.89

Table 2. Suppl. A summary of 10 candidate reference genes and 2 genes selected for data validation. Abbreviations used: bp- base pair, Ct- threshold cycle, SE- standard error, E- efficiency of PCR, F- forward, R- reverse.

Gene names	Gene descriptions	Accession numbers/locus IDs	Source Organisms	Primer sequences (5'-3')	Tm [°C]	Amplicon length [bp]	Ct	E [%]
NTB	<i>nucleotide tract-binding protein</i>	gi 242381788	<i>Phyllostachys edulis</i>	F-AGCTTATGGATATGTGGAGAA R-CTTCTTGGCGTGATTGATACT	55	111	26.15±0.26	104.93
		Os05g36120	<i>Oryza sativa</i>					
		Bradi2g24150	<i>Brachypodium distachyon</i>					
		TAE45871G003	<i>Triticum aestivum</i>					
		Zm00001d010173	<i>Zea mays</i>					
CAC	<i>clathrin adaptor complexes medium subunit</i>	Seita.3G228300	<i>Setaria italica</i>	F-TTTCTTCAGCAATGTGTGT R-CCCTTTTGCTTGTTTCTTGA	55	110	22.18±0.07	101.95
		gi 242375393	<i>P. edulis</i>					
		Os12g10560	<i>O. sativa</i>					
		Bradi4g40287	<i>B. distachyon</i>					
		TAE04243G001	<i>T. aestivum</i>					
CYP	<i>peptidyl prolyl cis-trans isomerase/cyclophilin</i>	Zm00001d007270	<i>Z. mays</i>	F-TTTACGGCGCCAAGTTCG R-TGGGAGCCGTTGGTGTG	55	100	25.86±0.21	106.61
		Seita.3G098000	<i>S. italica</i>					
		gi 242381616	<i>P. edulis</i>					
		Os09g39780	<i>O. sativa</i>					
		Bradi4g38880	<i>B. distachyon</i>					
18S	<i>18s ribosomal rna</i>	TAE04698G001	<i>T. aestivum</i>	F-GCAACTATGGTGCCTCTATG R-GCGACACCAGCAATCACCCA	55	105	24.38±0.12	105.16
		Zm00001d016660	<i>Z. mays</i>					
		Seita.2G137800	<i>S. italica</i>					
		gi 242375117	<i>P. edulis</i>					
		Os01g42480	<i>O. sativa</i>					
UBQ3	<i>polyubiquitin 3</i>	Bradi2g43200	<i>B. distachyon</i>	F-GAAGGAGTCCACCCTGCACCT R-CTGGATCTTGGCCTTCACGTT GTC	55	136	17.76±0.24	94.50
		TAE28272G001	<i>T. aestivum</i>					
		Zm00001d011325	<i>Z. mays</i>					
		Seita.5G222700	<i>S. italica</i>					
		AY954394.1	<i>O. sativa</i>					
ACT7	<i>actin 7</i>	PH01000093G1330	<i>P. edulis</i>	F-GGATTGGTGGTTCTATTTTGGC R-CACTTCATATGGACAATGCC	55	103	26.95±0.3	89.54
		Bradi3g04730	<i>B. distachyon</i>					
		TAE23060G001	<i>T. aestivum</i>					
		Zm00001d015327	<i>Z. mays</i>					
		Seita.4G259900	<i>S. italica</i>					
eEF1a	<i>eukaryotic elongation factor 1a</i>	XP_015641450	<i>O. sativa</i>	F-CGTGTGATTGAGAGGTTCGAGAA55 R- AGGTACCCGTGATCATGTTCT	55	211	22.15±0.32	94.06
		GU434145	<i>P. edulis</i>					
		Bradi2g56000	<i>B. distachyon</i>					
		TAE46567G001	<i>T. aestivum</i>					
		Zm00001d012277	<i>Z. mays</i>					
eIF4a	<i>eukaryotic initiation factor 4a</i>	Seita.5G393000	<i>S. italica</i>	F-ATGCTCTCCCGTGGTTTCAAG R-CAAGGGTAAGCTCATCTCTCTT CAC	55	172	22.92±0.31	91.42
		AK061464	<i>O. sativa</i>					
		PH01001781G0230	<i>P. edulis</i>					
		Bradi4g12750	<i>B. distachyon</i>					
		TAE11753G001	<i>T. aestivum</i>					
GAPDH	<i>glyceraldehyde-3-phosphate dehydrogenase</i>	Zm00001d037905	<i>Z. mays</i>	F-CCACAAACTGCCTTGCTCC R-CCCTGCCACCCCTCCAG	55	144	22.71±0.32	96.99
		Seita.3G265500	<i>S. italica</i>					
		XP_015626460	<i>O. sativa</i>					
		PH01002192G0160	<i>P. edulis</i>					
		Bradi1g34170	<i>B. distachyon</i>					
UBQ5	<i>ubiquitin 5</i>	TAE05656G001	<i>T. aestivum</i>	F-ATCACGCTGGAGGTGGAGT	55	213	22.32±0.22	109.47
		Zm00001d015251	<i>Z. mays</i>					
		Seita.1G089900	<i>S. italica</i>					
		XP_015650130	<i>O. sativa</i>					
		PH01003320G0070	<i>P. edulis</i>					

<i>TOC1</i>	<i>timing of cab expression1</i>	PH01001122G0410	<i>P. edulis</i>	R-CTTCTTCTTGCGCTTCTTGGC				
		Bradi2g34750	<i>B. distachyon</i>					
		TAE04445G002	<i>T. aestivum</i>					
		Zm00001d053838	<i>Z. mays</i>					
		Seita.7G270700	<i>S. italica</i>					
		KY249524	<i>Bambusa tulda</i>	F-AGGAACAAGGAACTGCGCCAC ATC	55	168	23.01±0.88	99.37
<i>MADS14</i>	<i>Mads14</i>	KX186751	<i>B. tulda</i>	R-GTCTTCGCCGCCACACATG F-TCGTGGAGAAGCAGAAGGTC R-CTGCTGCGTCCTCTGCCCT	55	163	24.97±0.59	95.03

Table 3. Suppl. The predicted stability ranking of 10 candidate reference genes using expression data obtained from 4 species (*Bambusa tulda*, *B. bambos*, *B. vulgaris*, and *B. balcooa*), 9 organs, and 3 developmental stages. This analysis was done using the *NormFinder* (NF) and *ReffFinder* (RF) programs. The values within *brackets* indicate the ranking order of the candidate reference genes in NF and RF analyses.

Species			<i>B. tulda</i> , <i>B. balcooa</i> , <i>B. bambos</i> , <i>B. vulgaris</i>			<i>B. tulda</i>	<i>B. tulda</i>		
Experimental sets			leaf	internode	rhizome	young leaves from flowering and non flowering culms, flag leaf, possible flag leaf, culm sheath, internode, root, rhizome, early staged inflorescence bud	leaf (tip, middle, base)	internode (tip, middle, base)	inflorescence bud (early, middle, late)
Reference genes	<i>ACT7</i>	NF	0.77 (6)	0.61 (6)	2.41 (10)	0.93 (7)	0.02 (1)	0.69 (10)	0.81 (10)
		RF	6.00 (6)	5.33 (5)	10.00 (10)	5.24 (5)	2.21 (1)	8.80 (10)	7.40 (10)
	<i>GAPDH</i>	NF	0.92 (7)	0.68 (8)	0.54 (2)	1.12 (9)	0.34 (6)	0.51 (6)	0.61 (8)
		RF	7.65 (7)	5.66 (6)	3.83 (4)	9.00 (10)	6.45 (7)	6.22 (8)	6.70 (8)
	<i>UBQ3</i>	NF	0.37 (4)	0.71 (9)	0.81 (5)	0.65 (4)	0.56 (9)	0.53 (7)	0.27 (9)
		RF	2.99 (4)	7.77 (9)	6.12 (8)	4.86 (4)	8.97 (10)	6.96 (9)	3.98 (4)
	<i>UBQ5</i>	NF	0.10 (2)	0.46 (4)	0.59 (3)	0.63 (3)	0.06 (2)	0.46 (4)	0.26 (2)
		RF	2.00 (2)	4.28 (4)	2.91 (3)	2.21 (2)	2.51 (2)	3.46 (4)	1.86 (1)
	<i>18S</i>	NF	1.07 (9)	0.67 (7)	0.98 (7)	1.31 (10)	0.65 (10)	0.64 (9)	0.63 (9)
		RF	8.45 (9)	7.04 (8)	4.92 (6)	5.62 (8)	5.62 (6)	5.20 (6)	5.20 (6)
	<i>CAC</i>	NF	0.10 (1)	0.13 (1)	1.07 (8)	0.89 (6)	0.26 (4)	0.61 (8)	0.32 (4)
		RF	1.57 (1)	1.41 (1)	4.76 (5)	5.29 (6)	2.63 (3)	5.66 (7)	4.00 (5)
	<i>CYP</i>	NF	1.45 (10)	0.59 (5)	0.75 (4)	0.54 (2)	0.30 (5)	0.46 (5)	0.13 (1)
		RF	9.74 (10)	5.73 (7)	2.38 (2)	4.05 (3)	2.66 (4)	5.01 (5)	2.78 (2)
	<i>NTB</i>	NF	0.96 (8)	0.77 (10)	0.28 (1)	0.80 (5)	0.45 (8)	0.25 (2)	0.56 (7)
		RF	8.00 (8)	9.15 (10)	1.50 (1)	5.32 (7)	7.67 (9)	2.78 (2)	7.27 (9)
	<i>eIF4α</i>	NF	0.30 (3)	0.33 (2)	0.94 (6)	0.54 (1)	0.40 (7)	0.14 (1)	0.44 (6)
		RF	2.45 (3)	2.00 (2)	5.96 (7)	1.41 (1)	7.45 (8)	2.00 (1)	5.42 (7)
	<i>eEF1α</i>	NF	0.47 (5)	0.44 (3)	1.36 (9)	0.95 (8)	0.18 (3)	0.34 (3)	0.44 (6)
		RF	4.40 (5)	2.91 (3)	9.00 (9)	6.62 (9)	4.21 (5)	2.82 (3)	3.64 (3)

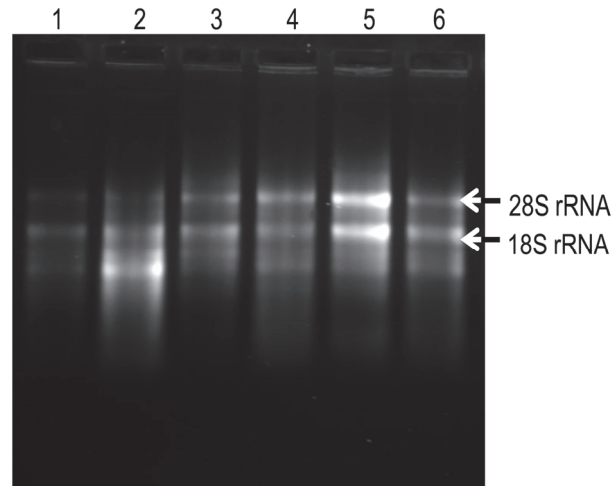


Fig. 1. Suppl. Assessment of quality of RNA isolated from a few of representative tissues used for this study. Representative agarose-formamide gel electrophoresis (1 %, m/v) of total RNA isolated from middle staged inflorescence bud (*lane 1*), root (*lane 2*), culm sheath (*lane 3*), rhizome (*lane 4*), leaf from non flowering culm (*lane 5*), and internode (*lane 6*) of *Bambusa tulda*.

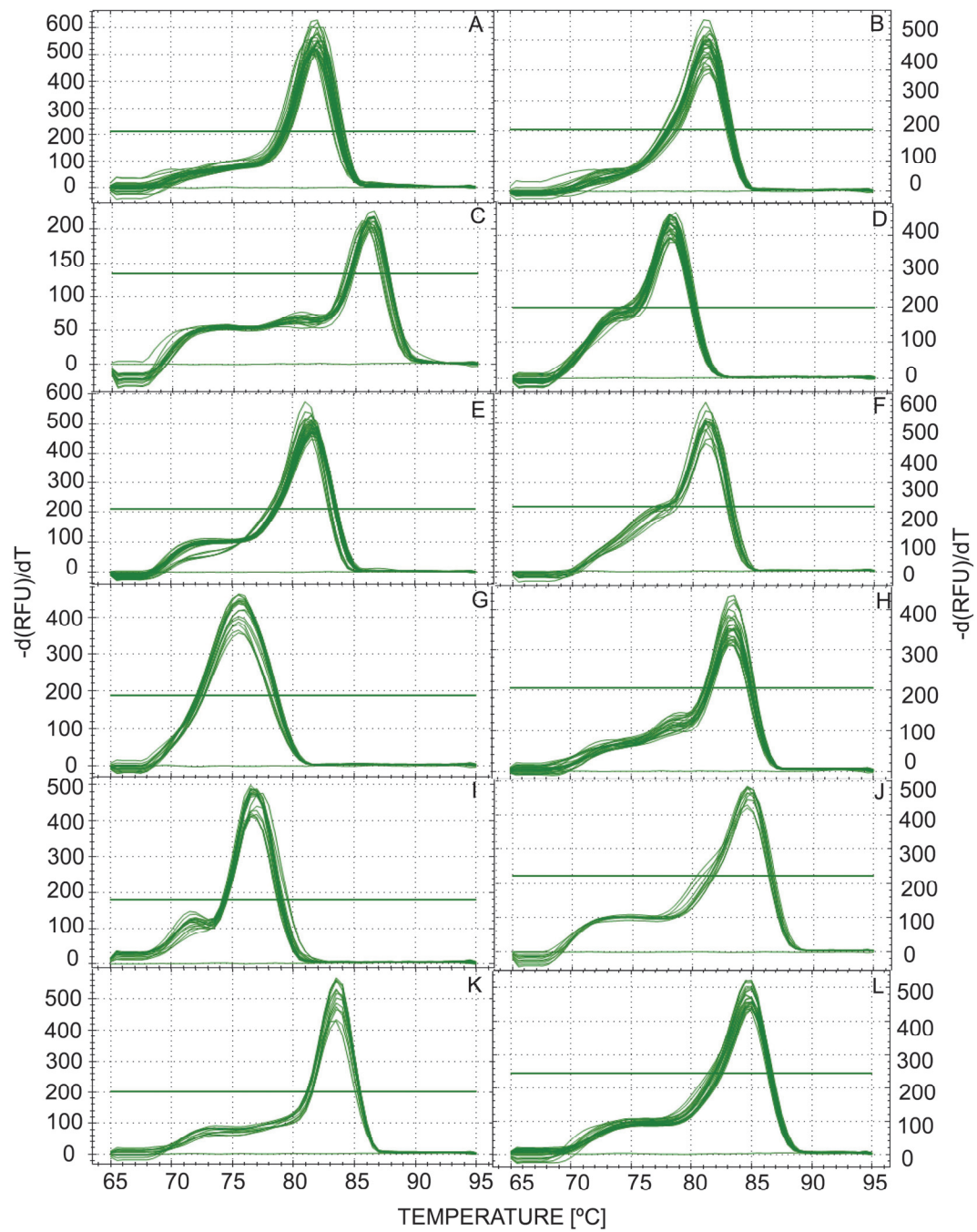


Fig. 2. Suppl. Melting curve analyses of 10 candidate reference genes *GAPDH* (A), *eIF4a* (B), *UBQ5* (C), *NTB* (D), *UBQ3* (E), *ACT7* (F), *CAC* (G), *eEF1a* (H), *CYP* (I), and *18S* (J), and two target genes *MADS14* (K) and *TOC1* (L) to verify primer specificity during real time PCR amplification. This analyses were performed in the *CFX* manager program (*Bio-Rad*, Hercules, USA) by observing the change in fluorescence as a consequence of increasing temperature [$-d(RFU)/dT$].

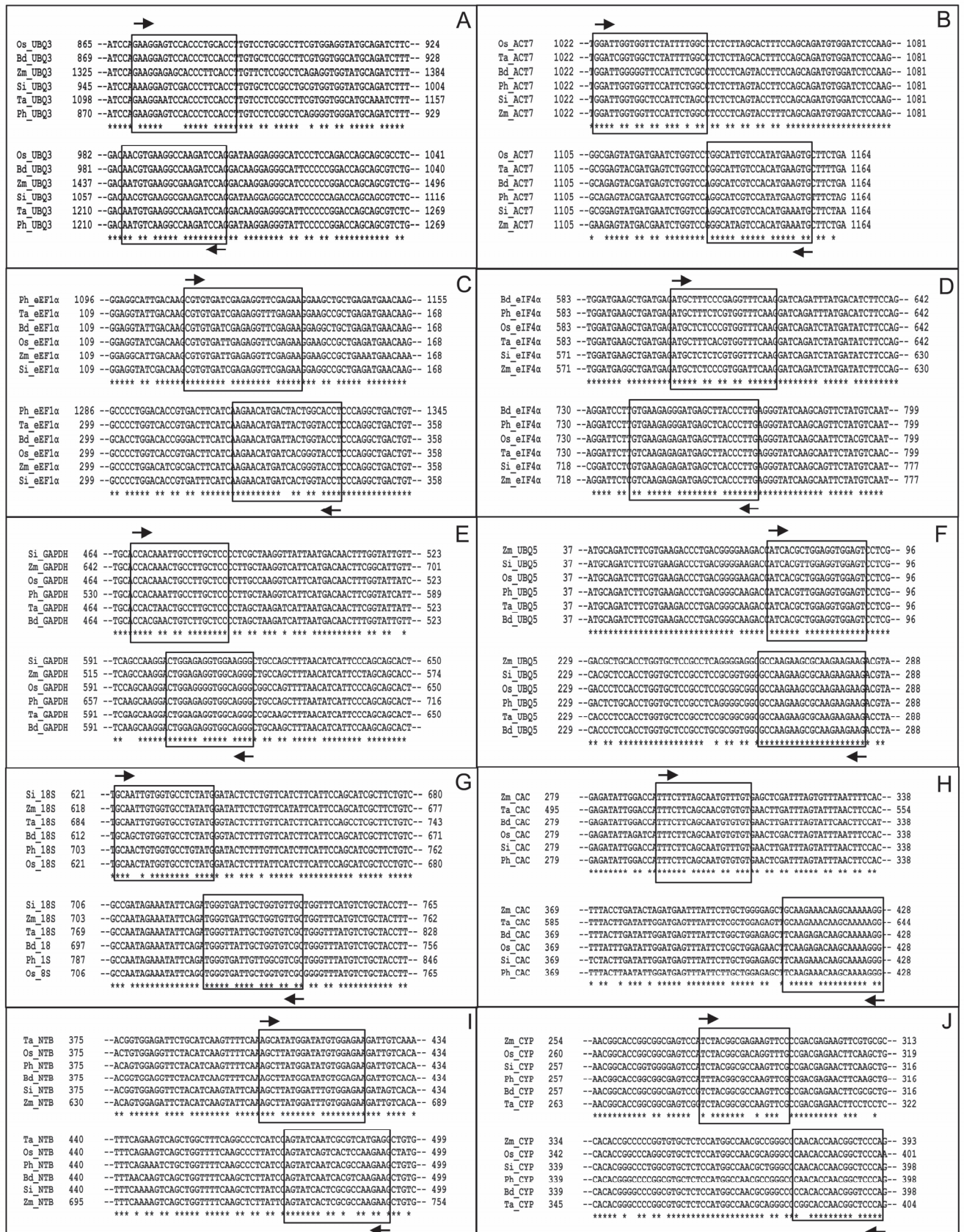


Fig. 3. Suppl. Multiple sequence alignment of homologous sequences of 10 candidate reference genes *UBQ3* (A), *ACT7* (B), *eEF1α* (C), *eIF4α* (D), *GAPDH* (E), *UBQ5* (F), *18S* (G), *CAC* (H), *NTB* (I), and *CYP* (J), obtained from sequenced monocot species. The *black boxes* indicate conserved regions used for primer designing. The *arrows* indicate direction of polymerization in PCR. *Os*- *Oryza sativa*, *Zm*- *Zea mays*, *Ta*- *Triticum aestivum*, *Si*- *Setaria italica*, *Bd*- *Brachypodium distachyon*, *Ph*- *Phyllostachys heterocycla*.

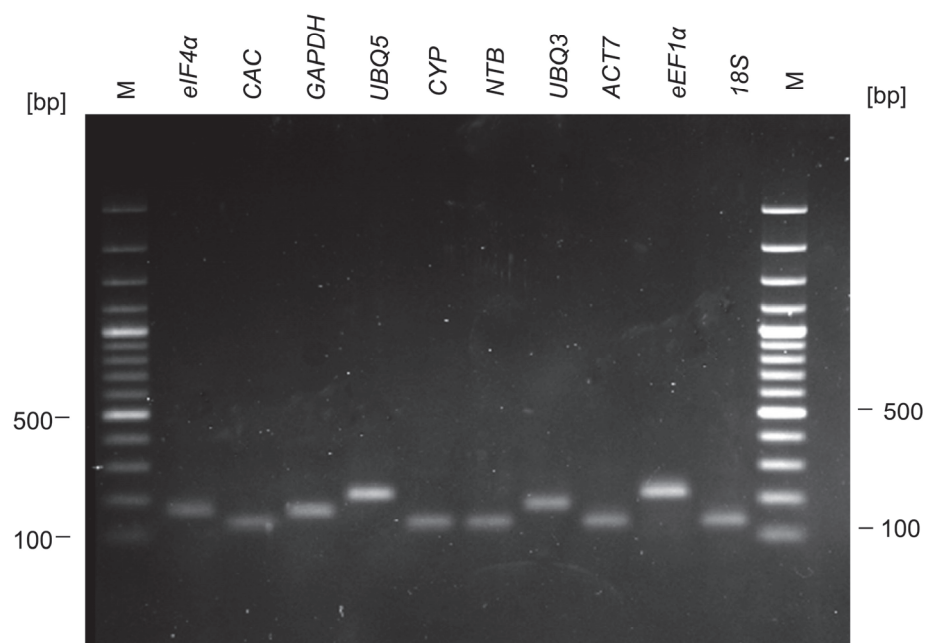


Fig. 4. Suppl. An agarose gel (2.5 %, m/v) showing a specific PCR amplification of selected 10 candidate reference genes of desired length from a cDNA library. M- molecular marker.