

Table 1 Suppl. The list of related genes and all primers of real time qPCR are shown below.

| Proteins | Gene abbreviation | Forward (F) and reverse (R) primers (5'-3') |
|--|-------------------|---|
| Chlorophyllide a oxygenase | <i>CAO</i> | F: TGGGTTTCAGTGAAAGAGGGT R: GAAGAGTGGCTGTAGGAGGA |
| Chlorophyll synthase | <i>CHLG</i> | F: ACCCACTTCGCTCTTCACA R: TTCTTGCTTGGCTCCTTTG |
| Magnesium-protoporphyrin IX monomethyl ester [oxidative] cyclase | <i>CRD1</i> | F: CAATGACTGGAAGGCAAAG R: ATCAAGGACAGCAGGGAAA |
| Magnesium-chelatase subunit ChII | <i>CHLI</i> | F: AGTCTGCTGCTTTATCCC R: GCAAGTTTCTGAGCCTGT |
| Protoporphyrinogen oxidase | <i>HEMG</i> | F: TATTCGTGATGAGCGTCTG R: TGTAGTTCAAGAGTAGCACCC |
| 5-Aminolevulinate dehydratase | <i>HEMB</i> | F: GGGTTGACTGATGAGGAGTG R: CTGACGACCTACGATTACGG |
| Porphobilinogen deaminase | <i>HEMC</i> | F: GAGTTGGAACACGAGGAA R: GAGGCTGACTTAATATCTTATC |
| Phytoene synthase | <i>PSY</i> | F: GGCTACACAACTGTTGAGAAC R: CCCAGGAGCTACTTTACCATATT |
| Lycopene β cyclase | <i>LYCB</i> | F: ATTGGTGGGAATTCAGGGATAG R: TTCAGCTAGTACTGGTGCTAAAG |
| Sedoheptulose-1,7-bisphosphatase | <i>SBPASE</i> | F: GCCACATTGACAATCCTG R: CATCGCTGCTGTAACTCC |
| Fructose-1,6-bisphosphatase | <i>FBPASE</i> | F: CATAGGTTGTCTCGTGGGT R: TTCGCTGATGTATCTGCTC |
| Photosystem I P700 apoprotein A1 | <i>PSAA</i> | F: AAACCCATTCTTACCACAA R: TAGCCTGTTTCCAATACTCA |
| Photosystem II protein D2 | <i>PSBD</i> | F: ACTCAAGCCGAAGAACT R: GCTAGACCGACTACTCCA |
| Photosystem II cytochrome b559 β subunit | <i>PSBF</i> | F: ATGACTATAGATCGAACCTATCCAA R: GCATTGCTGATATTGATCCCAA |
| Photosystem II phosphoprotein | <i>PSBH</i> | F: GGCTACACAACTGTTGAGAAC R: CCCAGGAGCTACTTTACCATATT |
| Superoxide dismutase [Cu-Zn] 2 | <i>Cu/Zn2-SOD</i> | F: TCATGCGGGTGATTAGG R: CTGCGTTTCCAGTTGTCTTA |
| Superoxide dismutase [Fe] | <i>Fe-SOD</i> | F: GTCATGTGCGAAGCCTAAA R: TAGCGTCAGTTCATCCAG |
| Cytosolic ascorbate peroxidase 2 | <i>APX2</i> | F: GCTCCTATTATGCTTCGTCT R: AGGTCCTCCCGTAACTTCA |
| Catalase 1 | <i>CAT1</i> | F: AGGGTAACTTTGATCTTGTT R: GTGTCTGTAATCCGTTGG |

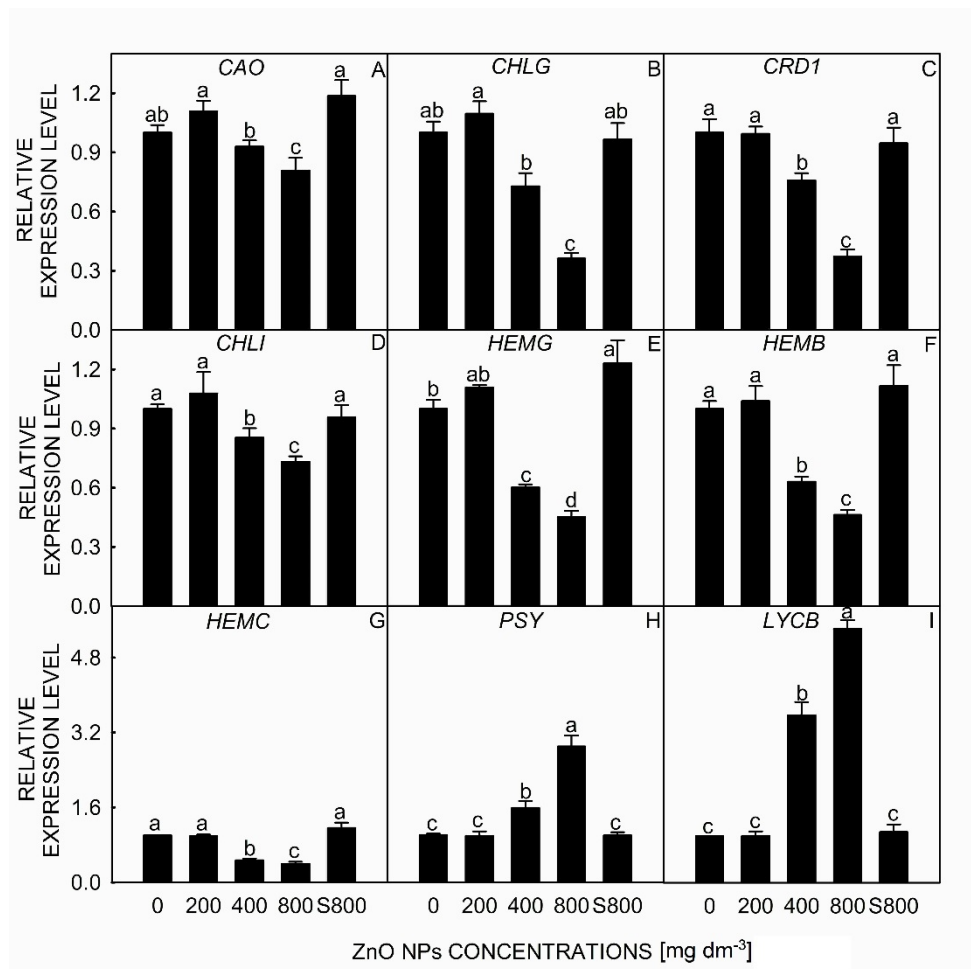


Fig. 1 Suppl. Effects of ZnO nanoparticles on the expression of genes connected with biosynthesis of photosynthetic pigments in tomato plants. RNA was isolated from leaves of plants treated with 0, 200, 400, and 800 mg dm⁻³ ZnO NPs, or supernatant from 800 mg dm⁻³ ZnO NP suspensions (S800). Real time qPCR was used to examine gene expressions; *ACT2* was used as a reference gene. The expressions of corresponding genes in tomato plants in the absence of ZnO NPs was set as 1. Means \pm SDs, $n = 4$; means with different letters are significantly different at 5 % level.

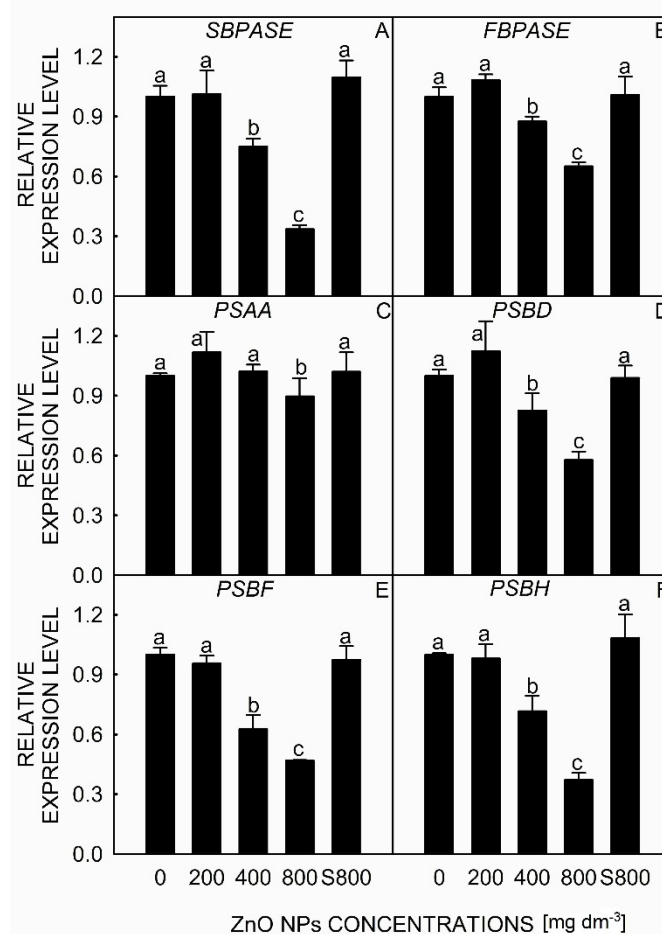


Fig. 2 Suppl. Effects of ZnO nanoparticles on the expression of genes connected with photosynthesis genes and photosystem structure in tomato plants. RNA was isolated from leaves of plants treated with 0, 200, 400, and 800 mg dm⁻³ ZnO NPs, or supernatant from 800 mg dm⁻³ ZnO NP suspensions (S800). Real time qPCR was used to examine the gene expressions; *ACT2* was used as a reference gene. The expressions of corresponding genes in the absence of ZnO NPs was set as 1. Means \pm SDs, $n = 4$; means with different letters are significantly different at 5 % level.

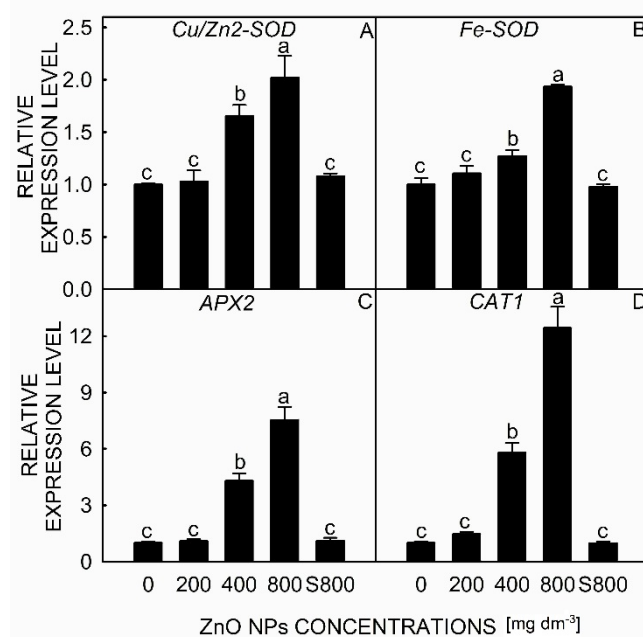


Fig. 3 Suppl. Effects of ZnO nanoparticles on the expression of genes encoding antioxidants in tomato plants. RNA was isolated from leaves of plants treated with 0, 200, 400, and 800 mg dm⁻³ ZnO NPs, or supernatant from 800 mg dm⁻³ ZnO NP suspensions (S800). Real time qPCR was used to examine the gene expressions and *ACT2* was used as a reference gene. The expressions of corresponding genes in tomato plants in the absence of ZnO NPs was set as 1. Means \pm SDs, $n = 4$; means with different letters are significantly different at 5 % level.