

Table 1 Suppl. The list of related genes and all primers of real time qPCR are shown below.

Proteins	Gene abbreviation	Forward (F) and reverse (R) primers (5'-3')
Chlorophyllide a oxygenase	<i>CAO</i>	F: TGGGTTCACTGAAAGAGGGT R: GAAGAGTGGCTGTAGGAGGA
Chlorophyll synthase	<i>CHLG</i>	F: ACCCACTTCGCTCTTCACA R: TTCTTGCTTGGCTCTTTG
Magnesium-protoporphyrin IX monomethyl ester [oxidative] cyclase	<i>CRD1</i>	F: CAATGACTGGAAGGCAAAG R: ATCAAGGACAGCAGGGAAA
Magnesium-chelatase subunit ChlI	<i>CHLI</i>	F: AGTCTGCTGCTTATCCC R: GCAAGTTCTGAGCCTGT
Protoporphyrinogen oxidase	<i>HEMG</i>	F: TATTCTGTATGAGCGTCTG R: TGTAGTTCAAGAGTAGCACCC
5-Aminolevulinate dehydratase	<i>HEMB</i>	F: GGGTTGACTGATGAGGAGTG R: CTGACGACCTACGATTACGG
Porphobilinogen deaminase	<i>HEMC</i>	F: GAGTTGGAACACGAGGAA R: GAGGCTGACTTAATATCTTATC
Phytoene synthase	<i>PSY</i>	F: GGCTACACAAACTGTTGAGAAC R: CCCAGGAGCTACTTACCATATT
Lycopene β cyclase	<i>LYCB</i>	F: ATTGGTGGATTCAAGGGATAG R: TTCAGCTAGTACTGGTGCTAAAG
Sedoheptulose-1,7-bisphosphatase	<i>SBPASE</i>	F: GCCACATTGACAATCCTG R: CATCGCTGCTGTAACCTCC
Fructose-1,6-bisphosphatase	<i>FBPASE</i>	F: CATAGGTTGTCTCGTGGGT R: TTCGCTGATGTATCTGCTC
Photosystem I P700 apoprotein A1	<i>PSAA</i>	F: AAACCCATTCTTACCAACAA R: TAGCCTGTTCCAATACTCA
Photosystem II protein D2	<i>PSBD</i>	F: ACTCAAGCCGAAGAAACT R: GCTAGACCGACTACTCCA
Photosystem II cytochrome b559 β subunit	<i>PSBF</i>	F: ATGACTATAGATCGAACCTATCCAA R: GCATTGCTGATATTGATCCAAA
Photosystem II phosphoprotein	<i>PSBH</i>	F: GGCTACACAAACTGTTGAGAAC R: CCCAGGAGCTACTTACCATATT
Superoxide dismutase [Cu-Zn] 2	<i>Cu/Zn2-SOD</i>	F: TCATGCGGGTGAATTAGG R: CTGCGTTCCAGTTGTCTTA
Superoxide dismutase [Fe]	<i>Fe-SOD</i>	F: GTCATGTCGAAGCCTAAA R: TAGCGTCAGTTCATCCAG
Cytosolic ascorbate peroxidase 2	<i>APX2</i>	F: GCTCCTATTATGCTTCGTCT R: AGGTCCCTCCGTAACTTCA
Catalase 1	<i>CAT1</i>	F: AGGGTAACTTGATCTTGTT R: GTGTCTGTAATCCGTTGG

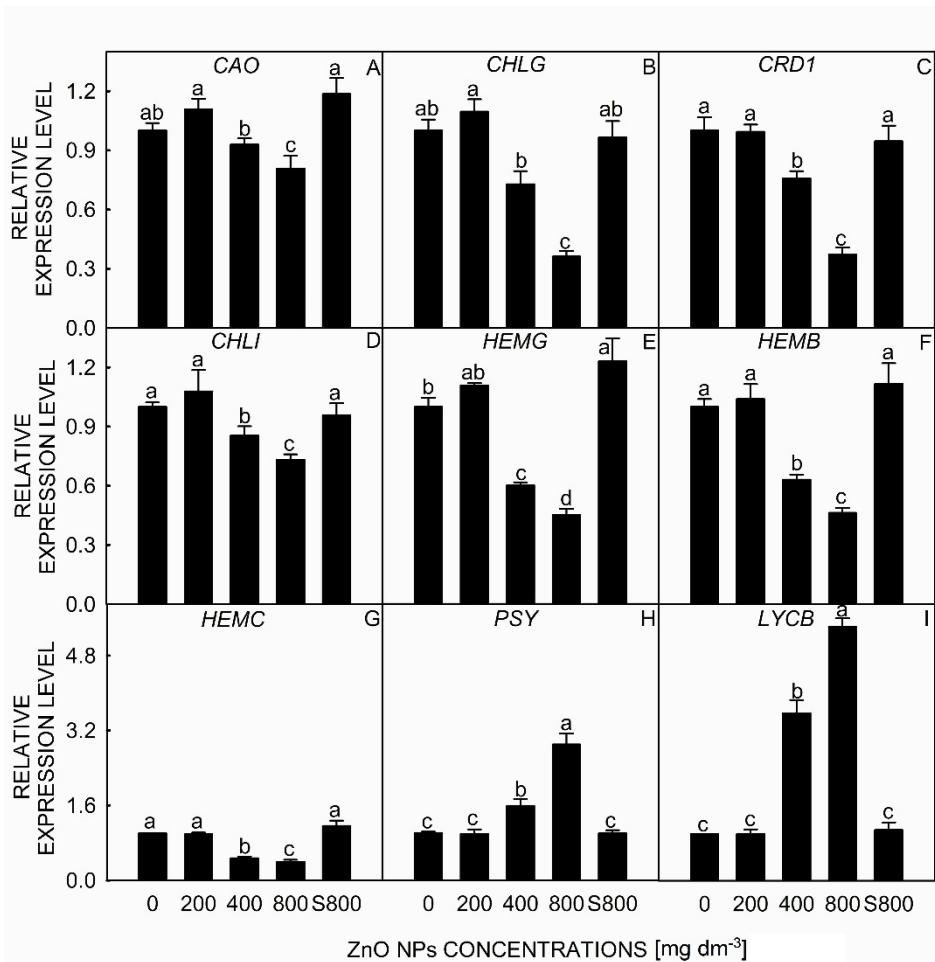


Fig. 1 Suppl. Effects of ZnO nanoparticles on the expression of genes connected with biosynthesis of photosynthetic pigments in tomato plants. RNA was isolated from leaves of plants treated with 0, 200, 400, and 800 mg dm^{-3} ZnO NPs, or supernatant from 800 mg dm^{-3} ZnO NP suspensions (S800). Real time qPCR was used to examine gene expressions; *ACT2* was used as a reference gene. The expressions of corresponding genes in tomato plants in the absence of ZnO NPs was set as 1. Means \pm SDs, $n = 4$; means with different letters are significantly different at 5 % level.

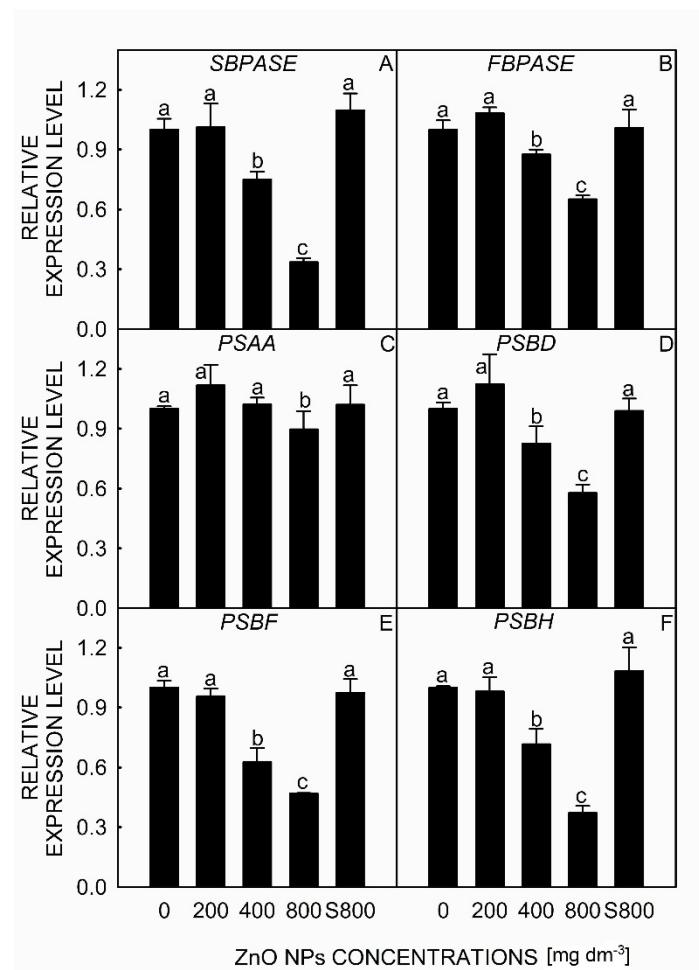


Fig. 2 Suppl. Effects of ZnO nanoparticles on the expression of genes connected with photosynthesis genes and photosystem structure in tomato plants. RNA was isolated from leaves of plants treated with 0, 200, 400, and 800 mg dm⁻³ ZnO NPs, or supernatant from 800 mg dm⁻³ ZnO NP suspensions (S800). Real time qPCR was used to examine the gene expressions; *ACT2* was used as a reference gene. The expressions of corresponding genes in the absence of ZnO NPs was set as 1. Means \pm SDs, $n = 4$; means with different letters are significantly different at 5 % level.

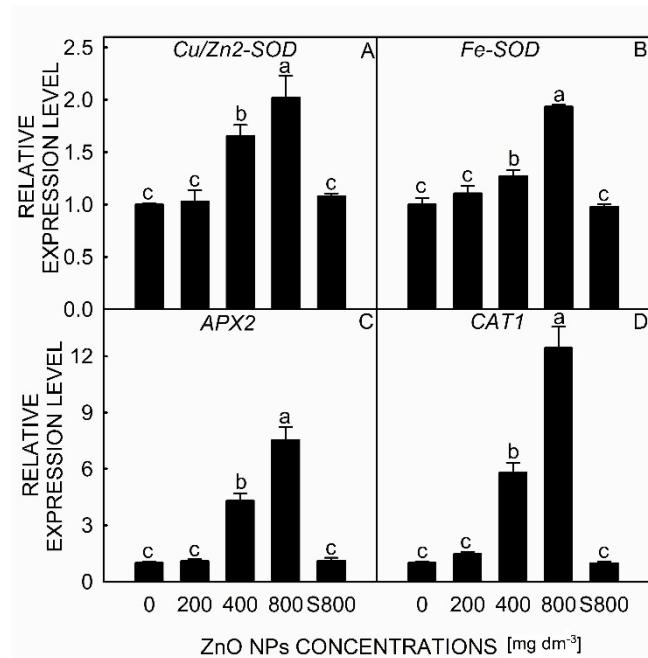


Fig. 3 Suppl. Effects of ZnO nanoparticles on the expression of genes encoding antioxidants in tomato plants. RNA was isolated from leaves of plants treated with 0, 200, 400, and 800 mg dm⁻³ ZnO NPs, or supernatant from 800 mg dm⁻³ ZnO NP suspensions (S800). Real time qPCR was used to examine the gene expressions and *ACT2* was used as a reference gene. The expressions of corresponding genes in tomato plants in the absence of ZnO NPs was set as 1. Means \pm SDs, $n = 4$; means with different letters are significantly different at 5 % level.