

Responses of *Nigella sativa* to foliar application of gibberellic acid and kinetin

S.H. SHAH¹, I. AHMAD and SAMIULLAH

Plant Physiology Section, Department of Botany, Aligarh Muslim University, Aligarh-202002, India

Abstract

Foliar sprays of water or 1, 10 and 100 μM aqueous solutions of gibberellic acid (GA_3) or kinetin (KIN) were applied to 40-d-old plants of *Nigella sativa* (L.) to study their effects on net photosynthetic rate, nitrogen metabolism, and the seed yield. 10 μM solutions of both the hormones, especially GA_3 , appreciably increased the activities of nitrate reductase and carbonic anhydrase, chlorophyll and total protein contents and net photosynthetic rate in the leaves, along with capsule number and seed yield plant⁻¹, at harvest.

Additional key words: black cumin, capsule number, carbonic anhydrase, leaf protein, net photosynthetic rate, nitrate reductase, seed yield.

Gibberellins (GAs) are a class of endogenous plant growth substances actively involved in the control of a number of key developmental processes including endosperm mobilization and stem elongation, as well as flower and fruit development (Hutty and Phillips 1995). Plants subjected to exogenous application of GAs have been found to exhibit increased activities of carbonic anhydrase (CA), nitrate reductase (NR) (Khan 1996, Hayat *et al.* 2001, Afroz *et al.* 2005), CO_2 fixation, stomatal conductance (Bishnoi *et al.* 1992), and ribulose-1,5-biphosphate carboxylase/oxygenase (RuBPCO) (Arteca and Dong 1981, Yuan and Xu 2001). Besides, GAs are also known to alter membrane permeability to ions (Manuel *et al.* 1980, Gilroy and Jones 1992), induce fruit set (Arteca 1996) and greatly enhance the translocation potential of the sink (Peretó and Beltrán 1987).

Despite the known importance of cytokinins (CKs) in plant growth and development, few studies have critically investigated the physiological impact of their exogenous application *e.g.*, increase in activities of nitrate reductase (Roa *et al.* 1984, Saxena and Saxena 2002), glutamate synthetase (Ghisi and Passera 1987), and enhancement of chlorophyll and protein contents (Towne and Owensby 1983, Synková *et al.* 1997). CKs may also be employed

to improve the photosynthetic rate and the activity of RuBPCO (Chernyad'ev 1994, Chernyad'ev and Mankhova 1998, Singh *et al.* 2001). Owing to the beneficence of mentioned physiological effects and the interwoven nature of actions of these hormones, we were tempted to study their impact on *Nigella sativa* L. in context of photosynthesis, nitrogen metabolism and yield.

Nigella sativa L. (black cumin) is a miraculously remedial herb, used intensely in medicinal as well as food formulations (Babayan *et al.* 1978, Takruri and Dameh 1998). Seeds of *Nigella sativa* L. were obtained from the Regional Research Institute of Unani Medicine, Aligarh (UP), India. They were surface sterilized with mercuric chloride solution (0.01 %), followed by repeated washings with double distilled water. The seeds were then sown in earthen pots filled with sandy loam soil and farmyard manure, mixed in a ratio of 9:1. A uniform basal dose (45, 300 and 78 mg) of N, P and K, in the form of urea, single superphosphate and muriate of potash, was applied at the time of sowing to each pot.

Gibberellic acid (GA_3) and kinetin (KIN) were obtained from Sigma Chemicals Co., St. Louis, USA. The plants of 40 d age were sprayed with 1, 10 or 100 μM GA_3 or KIN at the rate of 5 cm^3 plant⁻¹. Control plants were sprayed with double distilled water only.

Received 14 March 2006, accepted 20 July 2006.

Abbreviations: CA - carbonic anhydrase; Chl - chlorophyll; CK - cytokinin; DAS - days after sowing; GAs - gibberellins; GA_3 - gibberellic acid; KIN - kinetin; NR - nitrate reductase; P_{N} - net photosynthetic rate.

Acknowledgement: The authors wish to thank Prof. A. Ridhal for his critical reading of the manuscript and valuable comments.

¹ Corresponding author, fax: (+91) 571 2702885; e-mail: shoukathusseine@gmail.com

Each treatment was replicated three times. The pots were lined in the Department's net house, according to simple randomized block design. The various parameters were studied at 50, 70 and 90 d after spray.

Carbonic anhydrase (E.C. 4.2.1.1) activity was assayed by the procedure adopted by Dwivedi and Randhawa (1974). 200 mg fresh leaf was cut into small pieces in 0.2 M cysteine, at 4 °C. These pieces were transferred to test tube with phosphate buffer and 0.2 M sodium bicarbonate and bromothymol blue were added. CO₂ liberated during catalytic action of enzyme on NaHCO₃ was estimated by titrating the reaction mixture against 0.01 M HCl, using methyl red as an indicator. Nitrate reductase (E.C. 1.6.6.1) activity was determined in fresh leaves of the plants by the method of Jaworski (1971). Total chlorophyll content was estimated following the method of Mackinney (1941). Net photosynthetic rate (P_N) at each stage of sampling was measured in fully expanded leaves by a portable photosynthetic system (LI-COR 6200, Lincoln, NE, USA). Protein content in the leaves was estimated by the method of Lowry *et al.* (1951). Treatment means were compared by analysis of variance using statistical package SPSS (SPSS 7.5.1 for Windows, standard version 1996). Least significant difference (LSD) was estimated at 0.05 level of probability.

Although, spray of either GA₃ or KIN was found to promote activity of CA, GA₃ was more efficient with 45 % increase in CA activity (relative to control) in 70-d old plants when sprayed with 10 µM GA₃ at 40-d-stage (Table 1). Enzyme activity perceptibly increased with the increase in hormone concentration from 1 to 10 µM, however, a higher concentration, *i.e.*, 100 µM of either KIN or GA₃, failed to bring about any appreciable stimulation. Maybe KIN induced an increase in endogenous auxin content to supraoptimal level, thereby initiating an inhibitory regulation of its own biosynthesis and action (Kamínek *et al.* 1997). Similarly, GAs are known to check their own effects through feedback control under excess concentrations (Martin *et al.* 1996). The observed increase in CA activity induced by 1 to 10 µM GA₃ can be attributed to its effect on the *de novo* synthesis of CA involving the translational and transcriptional mechanisms (Okabe *et al.* 1980, Khan *et al.* 2004). Similarly, the mode of action of KIN can be traced to the influence of cytokinins at the level of transcription and/or stabilization of the transcripts as they increased the content of CA-mRNA (Sugiharto *et al.* 1992).

In context to the NR activity, GA₃ again outperformed KIN as better inducer of such enzyme activity (Table 1). The positive effect of GAs on NR activity is well known (Khan 1996, Premabatidevi 1998). A marked rise in the protein content (Table 1) of the treated leaves may well be a consequence of this GA₃ intensified NR activity (Saroor *et al.* 1998). This postulation is further complemented by the positive correlation between the content of proteins and that of the enzyme ($r = 0.905^{**}$). Here also, the influences of KIN and GA₃ on the

translation/transcription mechanisms, as mentioned above, may be stated in explanation for the increase in NR activity and protein content.

Leaves of the plants receiving KIN treatment, had higher chlorophyll content compared to control (Table 1),

Table 1. Carbonic anhydrase (CA) activity [mol(CO₂) kg⁻¹ s⁻¹], nitrate reductase (NR) activity [nmol(NO₂) g⁻¹ min⁻¹], chlorophyll (Chl) [g kg⁻¹(f.m.)] and protein content [% (d.m.)], and net photosynthetic rate (P_N) [µmol(CO₂) m⁻² s⁻¹] in *Nigella sativa* leaves, sprayed with water (control), kinetin (KIN), or gibberellic acid (GA₃) at 40 d after sowing and sampled at 50, 70 and 90 DAS. LSD for $P = 0.05$, mean \pm SE.

Parameter	Treatment	[µM]	50 DAS	70 DAS	90 DAS
CA	control		2.07 \pm 0.18	2.54 \pm 0.19	1.35 \pm 0.12
	KIN	1	2.41 \pm 0.21	3.02 \pm 0.22	1.65 \pm 0.14
		10	2.68 \pm 0.19	3.43 \pm 0.26	1.95 \pm 0.20
		100	2.56 \pm 0.23	3.34 \pm 0.23	1.85 \pm 0.15
	GA ₃	1	2.51 \pm 0.22	3.15 \pm 0.25	1.75 \pm 0.14
		10	2.87 \pm 0.29	3.68 \pm 0.27	2.15 \pm 0.21
		100	2.65 \pm 0.19	3.25 \pm 0.24	1.87 \pm 0.16
	LSD		0.071	0.051	0.041
	NR	control	7.04 \pm 0.72	7.66 \pm 0.84	4.88 \pm 0.91
		KIN	1	7.42 \pm 0.59	8.52 \pm 0.57
			10	7.87 \pm 0.63	9.52 \pm 0.71
			100	8.03 \pm 0.89	9.50 \pm 0.82
		GA ₃	1	7.64 \pm 0.61	8.65 \pm 0.72
			10	8.67 \pm 0.67	9.92 \pm 0.76
			100	8.09 \pm 0.74	9.42 \pm 0.87
		LSD		0.41	0.75
					0.31
Chl	control		1.24 \pm 0.11	1.26 \pm 0.11	0.97 \pm 0.09
		KIN	1	1.36 \pm 0.12	1.38 \pm 0.12
			10	1.48 \pm 0.11	1.57 \pm 0.15
			100	1.50 \pm 0.12	1.54 \pm 0.14
		GA ₃	1	1.42 \pm 0.13	1.46 \pm 0.11
			10	1.62 \pm 0.15	1.68 \pm 0.17
			100	1.52 \pm 0.14	1.53 \pm 0.14
		LSD		0.07	0.08
					0.05
	Protein	control	11.40 \pm 1.2	12.35 \pm 1.2	10.40 \pm 1.1
		KIN	1	13.01 \pm 1.1	14.74 \pm 1.3
			10	14.65 \pm 1.3	17.60 \pm 1.4
			100	14.80 \pm 1.5	16.80 \pm 1.3
		GA ₃	1	12.95 \pm 1.1	15.06 \pm 1.4
			10	15.41 \pm 1.4	18.70 \pm 1.7
			100	14.21 \pm 1.2	17.71 \pm 1.6
		LSD		0.71	0.52
					0.65
P _N	control		15.10 \pm 1.4	16.10 \pm 1.5	13.25 \pm 1.3
		KIN	1	16.21 \pm 1.5	18.41 \pm 1.8
			10	17.29 \pm 1.6	19.75 \pm 1.7
			100	16.85 \pm 1.4	20.01 \pm 1.4
		GA ₃	1	16.35 \pm 1.3	19.25 \pm 1.5
			10	19.01 \pm 1.7	22.15 \pm 1.9
			100	17.15 \pm 1.6	20.45 \pm 1.4
		LSD		0.65	0.46
					0.75

which is in agreement with Ghosh and Biswas (1991), Saroop *et al.* (1994) and Saxena and Saxena (2002). However, such an effect of GA₃ was more pronounced than that of KIN (Table 1), which may be attributed to the GA₃-generated enhancement of ultrastructural morphogenesis of plastids, coupled with the retention of Chl and delay of senescence caused by the hormone treatment (Arteca 1996, Ouzounidou and Illias 2005), and the increase in protein content. Similarly, CKs can also be said to have promoted Chl content through conversion of etioplasts into chloroplasts, and by governing the composition and ultrastructure of plastids (Pospíšilová *et al.* 2000). A subsequent expression of the cumulative effect of increased activities of NR and CA, and protein and Chl contents was the high P_N rate (Table 1) of the hormone (GA₃ or KIN) treated plants. Various other

Table 2. Number of capsules and seed yield plant⁻¹ in *Nigella sativa* L. plants, sprayed with water (control), gibberellic acid (GA₃) or kinetin (KIN) at 40 d after sowing and sampled, at harvest (130 DAS), LSD for $P = 0.05$, mean \pm SE.

Treatment	[μM]	Number of capsules [plant ⁻¹]	Seed yield [g plant ⁻¹]
Control		16.20 \pm 1.5	1.12 \pm 0.16
KIN	1	18.35 \pm 1.8	1.25 \pm 0.19
	10	21.50 \pm 1.9	1.50 \pm 0.22
	100	20.91 \pm 1.4	1.41 \pm 0.24
GA ₃	1	19.70 \pm 1.5	1.32 \pm 0.18
	10	23.75 \pm 2.1	1.72 \pm 0.19
	100	21.52 \pm 1.9	1.52 \pm 0.20
LSD		1.02	0.08

References

Afroz, S., Mohammad, F., Hayat, S., Siddiqui, M.H.: Exogeneous application of gibberellic acid counteracts the ill effects of sodium chloride in mustard. - Turk. J. Biol. **29**: 233-236, 2005.

Arteca, R.N.: Plant Growth Substances: Principles and Applications. - CBS Publishers, New Delhi 1996.

Arteca, R.N., Dong, C.N.: Stimulation of photosynthesis by application of phytohormones to root systems of tomato plants. - Photosynth. Res. **2**: 243-249, 1981.

Azam, Z.M.: Response of *Plantago ovata* and *Trigonella foenum-graecum* to N, P and GA₃ application. - Ph.D. Thesis, Aligarh Muslim University, Aligarh 2002.

Babayan, V.K., Koottungal, D., Halaby, G.A.: Proximate analysis, fatty acid and amino acid composition of *Nigella sativa* L. seeds. - J. Food Sci. **43**: 1314-1315, 1978.

Bishnoi, N.R., Krishnamoorthy, H.N.: Effect of waterlogging and gibberellic acid on leaf gas-exchange in peanut (*Arachis hypogaea* L.). - J. Plant Physiol. **139**: 503-505, 1992.

Bruinsma, J.: Rolle der Cytokinine bei Blüten und Furchtentwicklung. - Z. Pflanzenernähr. Boden **140**: 15-23, 1977.

Chernyad'ev, I.I., Monakhova, O.F.: The activity and content of ribulose-1,5-biphosphate carboxylase/oxygenase in wheat plants as affected by water stress and kartolin-4. - Photosynthetica **35**: 603-610, 1998.

Chrungoo, N.K., Farooq, S.: Influence of gibberellic acid and naphthalene acetic acid on yield and growth of saffron (*Crocus sativus* L.). - Indian J. Plant Physiol. **27**: 201-205, 1984.

Dwivedi, R.S., Randhawa, N.S.: Evaluation of a rapid test for the hidden hunger of zinc in plants. - Plant Soil **40**: 445-451, 1974.

Ghisi, R., Passera, C.: Effect of benzylaminopurine on enzyme activities of nitrogen assimilation pathway in detached leaves of maize plants. - Agrochimica **31**: 265-272, 1987.

Ghosh, A.K., Biswas, A.K.: Source-sink relationship during ageing and senescence of *Solanum tuberosum* L. - Indian J. Plant Physiol. **34**: 25-29, 1991.

Gilroy, S., Jones, R.L.: Gibberellic acid and abscisic acid coordinately regulate cytoplasmic calcium and secretory activities in barley aleurone protoplast. - Proc. nat. Acad. Sci. USA **89**: 3591-3595, 1992.

Hayat, S., Ahmad, A., Mobin, M., Fariduddin, Q., Azam, Z.M.: Carbonic anhydrase, photosynthesis and seed yield in mustard plants treated with phytohormones. -

factors such as, ample availability of CO₂ due to increased CAactivity, along with GA₃ induced promotion of the rates of cyclic and non-cyclic phosphorylation (Saroop *et al.* 1994, Naidu and Swamy 1995) and the activity of RuBPCO, may also have supplemented the above stance. Indeed, the activity of CA was positively correlated with P_N ($r = 0.831^{**}$).

Vegetative growth of a test plant is highly responsible for desired seed yield, as the number of photosynthesizing sites is detrimental in the production of photoassimilates. The rise in photosynthetic rate implies more vegetative growth due to sufficient availability of nutrients, which in turn, increases the number and size of the reproductive sink, *i.e.*, number of flowers and capsules (Table 2). This, when coupled with the GA₃ induced mobilization of metabolites to the developing pods (Hoad *et al.* 1977, Naidu and Swamy 1995, Ouzounidou and Illias 2005), may have stimulated their growth and hence, caused an increase in seed yield, as observed herein (Table 2). Similary, KIN was also seen to have some effect on yield, though relatively subdued, which may be ascribed to the increased opportunity for formation of new buds (Bruinsma 1977), and successful reversal of leaf and fruit (pod) abscission . Such favourable effect of KIN or GA₃ in enhancing economic yield of *Nigella sativa* and various other crops has also been reported by Chrungoo and Farooq (1984), Khan (1996), Hayat *et al.* (2001), Mousa *et al.* (2001), and Azam (2002). Summarily, we can say, that the observed increase in seed yield is therefore, an expression of the cumulative effect of the hormones on the leaf metabolism, re-directed mobili-zation of metabolites, and delayed senescence of plant organs.

Photosynthetica **39**: 111-114, 2001.

Hoad, G.V., Loveys, B.R., Skenek, G.M.: The effect of fruit removal on cytokinins and gibberellin-like substances. - *Planta* **136**: 25-30, 1977.

Hutty, A.K., Phillips, A.L.: Gibberellin regulated plant genes. - *Physiol. Plant.* **95**: 310-317, 1995.

Jaworski, E.G.: Nitrate reductase assay in intact plant tissue. - *Biochem. biophys. Res. Commun.* **43**: 1274-1279, 1971.

Kamínek, M., Motyka, V., Vaňková, R.: Regulation of cytokinin content in plant cells. - *Physiol. Plant.* **101**: 689-700, 1997.

Khan, N.A.: Effect of gibberellic acid on carbonic anhydrase, photosynthesis, growth and yield of mustard. - *Biol. Plant.* **38**: 145-147, 1996.

Khan, N.A., Javid, S., Samiullah: Physiological role of carbonic anhydrase in CO_2 fixation and carbon partitioning. - *Physiol. mol. Biol. Plants* **10**: 153-166, 2004.

Lowry, O.H., Rosebrough, N.J., Farr, A.L., Randall, R.J.: Protein measurement with Folin-phenol reagent. - *J. biol. Chem.* **193**: 265-275, 1951.

Mackinney, G.: Absorption of light by chlorophyll solutions. - *J. biol. Chem.* **140**: 315-322, 1941.

Manuel, D., De la Guardia, M.B.: Effects of potassium and gibberellic acid on stem growth of whole sunflower plants. - *Physiol. Plant.* **49**: 443-448, 1980.

Martin, D.N., Proebsting, W.M., Parks, T.D., Dougherty, W.G., Lange, T., Lewis, M.J., Gaskin, P., Heddin, P.: Feedback regulation of gibberellin metabolism and gene expression in *Pisum sativum* L. - *Planta* **200**: 159-166, 1996.

Mousa, G.T., El-Sallami, I.H., Ali, E.F.: Response of *Nigella sativa* L. to foliar application of gibberellic acid, benzyladenine, iron and zinc. - *Assiut J. agr. Sci.* **32**: 141-156, 2001.

Naidu, C.V., Swamy, P.M.: Effect of gibberellic acid on growth biomass production and associated physiological parameters in some selected tree species. - *Indian J. Plant Physiol.* **38**: 15-17, 1995.

Okabe, K., Lindlar, A., Tsuzuki, M., Miyachi, S.: Carbonic anhydrase on ribulose 1,5-biphosphate carboxylase and oxygenase. - *FEBS Lett.* **114**: 142-144, 1980.

Ouzounidou, G., Ilias, I.: Hormone-induced protection of sunflower photosynthetic apparatus against copper toxicity. - *Biol. Plant.* **49**: 223-228, 2005.

Peretó, J.G., Beltrán, J.P.: Hormone directed sucrose transport during fruit set induced by gibberellins in *Pisum sativum*. - *Physiol. Plant.* **69**: 356-360, 1987.

Pospíšilová, J., Synková, H., Rulcová, J.: Cytokinins and water stress. - *Biol. Plant.* **43**: 321-328, 2000.

Premabatidevi, R.K.: Effect of IAA, GA_3 , and kinetin on nitrate reductase and nitrite reductase in the leaves of a tree legume. - *Indian J. Plant Physiol.* **3**: 97-101, 1998.

Roa, L.V.M., Datta, N., Mahadevan, M., Guha-Mukherjee, S., Sopory, S.S.: Influence of cytokinins and phytochrome on nitrate reductase activity in etiolated leaves of maize. - *Photochemistry* **23**: 1875-1879, 1984.

Saroop, S., Thaker, V.S., Chanda, S.V., Singh, Y.D.: Light and nitrate induction of nitrate reduction in kinetin and gibberellic acid treated mustard cotyledons. - *Acta Physiol. Plant.* **20**: 359-362, 1998.

Saroop, S., Thaker, V.S., Chanda, S.V., Vaishnav, P.P.: Development of photosynthetic electron transport reactions under the influence of phytohormones during greening of cotyledons of *Brassica juncea*. - *Photosynthetica* **30**: 193-200, 1994.

Saxena, A., Saxena, D.K.: Nitrate reductase activity and chlorophyll content in *Sphagnum* as affected by kinetin, glutathione and metals. - *Indian J. Plant Physiol.* **7**: 83-85, 2002.

Singh, D.V., Srivastava, G.C., Abdin, M.Z.: Amelioration of negative effect of water stress in *Cassia angustifolia* by benzyladenine and/or ascorbic acid. - *Biol. Plant.* **44**: 141-143, 2001.

Sugiharto, B., Sugiyama, T.: Effects of nitrate and ammonium on gene expression of phosphoenolpyruvate carboxylase and nitrogen metabolism in maize leaf tissue during recovery from nitrogen stress. - *Plant Physiol.* **98**: 1403-1408, 1992.

Synková, H., Wilhelmová, N., Šesták, Z., Pospíšilová, J.: Photosynthesis in transgenic plants with elevated cytokinin contents. - In: Pessarakli, M. (ed.): *Handbook of Photosynthesis*. Pp. 541-552. Marcel Dekker, New York - Basel - Hong Kong 1997.

Takruri, H.R.H., Dameh, M.A.F.: Study of the nutritional value of black cumin seeds (*Nigella sativa* L.). - *J. Sci. Food Agr.* **76**: 404-410, 1998.

Towne, G., Owensesby, C.: Cytokinins effect on protein and chlorophyll content of big bluestem leaves. - *J. Range Manage.* **36**: 75-77, 1983.

Yuan, L., Xu, D.Q.: Stimulation effect of gibberellic acid short-term treatment on leaf photosynthesis related to the increase in Rubisco content in broad bean and soybean. - *Photosynth. Res.* **68**: 39-47, 2001.